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Rapid and efficient microwave-assisted synthesis of highly sulfated organic scaffolds

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Abstract—Sulfation of multiple hydroxylated small organic molecules is fraught with problems of poor yield, multitude of products, and long reaction times. We have developed a rapid microwave-based method for synthesis of highly sulfated small organic molecules, which affords the per-sulfated product in moderate to excellent yields and high purity. The method is expected to be of value in the discovery of per-sulfated organic molecules as mimics of glycosaminoglycans, which are being increasingly recognized as modulators of key physiological functions.
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Recent work in our laboratory shows that designed highly sulfated, aromatic, small organic molecules possess interesting physico-chemical and biological properties.[1–5] Biochemically, these molecules form multiple ionic as well as non-ionic interactions, which form the backbone of most protein-recognition elements. Structurally, these represent mimics of glycosaminoglycans (GAGs), which are increasingly being recognized as modulators of key physiological functions,[6,7] while toxicologically, the sulfated structure represents a highly water-soluble, already-metabolized form that is expected to possess minimal toxicity. Despite these novel features, highly sulfated organic molecules remain largely unexplored.

A major limitation in exploring these novel structures is their challenging synthesis. Nearly all small organic sulfates reported in the literature are mono- or di-sulfated molecules,[8–12] typically prepared using sulfur trioxide complexes with amines in a highly polar solvent (DMF or DMA). Sulfation of such organic scaffolds may require as many as 13 h and temperatures as high as 95 °C in the presence of a large excess of the sulfating complexes,[8–12] while sugars, which contain multiple –OH groups, require reaction times in the range of 12 h to several days.[13–19]

Theoretically, this method could be extended for the synthesis of highly sulfated drug-like molecules, yet practically it is a synthetic nightmare because these molecules possess significantly higher negative charge density.[2] The major challenge is driving the reaction to completion in order to sulfate all available hydroxyl groups (alcoholic or phenolic) on the substrate. As the number of –OH groups increase on a small scaffold, sulfation becomes progressively difficult because of anion crowding, resulting in numerous partially sulfated side-products.

A further challenge is the isolation of the chemically pure per-sulfated product, which requires aqueous isolation techniques. Yields in the range of 11% and 100% have been reported, yet the presence of inorganic salts arising from the use of buffers and salts lead to significant inconsistencies and inaccuracies. Additionally, instability of highly anionic products introduce limitations on reaction times and temperature.[13] This is likely to be especially true for highly sulfated, aromatic, small organic molecules, which are expected to be less stable than the saccharide scaffolds.

To avoid these problems with one-step sulfation, we recently synthesized some small aromatic per-sulfated structures using a two-step approach involving the 2,2,2-trichloroethyl protecting group.[20] The two-step protection–deprotection protocol resolved some of the

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problems of the direct sulfation approach, yet required careful real-time monitoring of the reaction by RP-HPLC to prevent product degradation and was not particularly applicable to substrates that were acid and/or metal sensitive. These limitations led us to seek an alternative sulfation approach, which can be rapid, efficient, and widely applicable to a number of poly-hydroxy scaffolds.

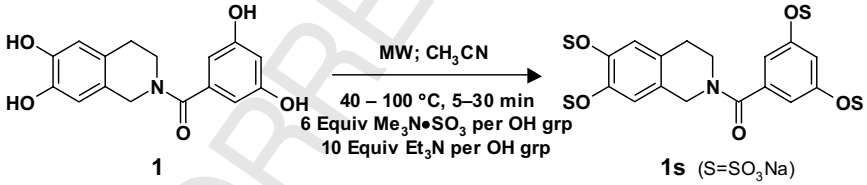
We hypothesized that significant rate enhancements are likely to be achieved using microwaves, especially because the ionic sulfated product may couple to microwaves through ionic conduction, for example, in CH_3CN.[21] CH_3CN was chosen as the solvent over the commonly used DMF because (a) it can be evaporated at lower temperature (thus aiding isolation) and (b) it was likely to solubilize the per-sulfated product with an amine counter-ion. We also hypothesized that introducing free base in the reaction mixture should promote the difficult per-sulfation reaction.

Sulfation of **1**[22] with $SO_3{\cdot}Me_3N$ complex (6 equiv per –OH group) at 100 °C in the absence of free base gave only 4.7% of per-sulfated product **1s** in 20 min (Table 1, entry 1).[23] Inclusion of 1 equiv of free Et_3N per –OH group resulted in 13.5% conversion (entry 2), while 79.8% of **1s** was formed with 5 equiv of Et_3N (entry 3). Further increase to 10 equiv Et_3N per –OH group had a negligible increase in the yield of **1s** (entry 4). Increasing the proportion of $SO_3{\cdot}Me_3N$ per –OH group from 1 to 9 M equiv (Table 1, entries 5–8) gradually increased the yield of the per-sulfated product from 14.5% to 79.5%, while further increase to 12 equiv was found to be not particularly advantageous (entry 9). For further studies, 6 and 10 M equiv of the sulfating complex and base, respectively, were chosen.

To assess the effect of solvent, we chose to evaluate nitromethane and DMF, both of which are solvents with high dielectric constant and known to be microwave-friendly. While only 23.2% and 17.2% of per-sulfated product **1s** was formed from **1** in 10 min at 100 °C in CH_3NO_2 and DMF, respectively, 47.4% of the product was formed in CH_3CN (Table 1, entries 10 and 11). Thus, our initial choice of CH_3CN proved to be optimal. To assess the effect of temperature and reaction time, sulfation was performed for 10–30 min at 40 to 120 °C. While 30 min were required to yield 91.2% of **1s** at 100 °C, only 10 min were needed for 90.8% conversion at 120 °C. In striking contrast, no product was detected at 40 °C within 10 min. Finally, $SO_3{\cdot}$py/py was found to give nearly twice as much per-sulfated product as $SO_3{\cdot}Me_3N/Et_3N$ (entries 7 and 12) in 10 min at 100 °C. Since pyridine is ~10,000-fold weaker base in comparison to Et_3N, this result suggests general base catalysis as the predominant mechanism of sulfation rather than a process involving deprotonation of the substrate followed by nucleophilic attack.

Appropriate control reactions in the absence of microwaves using two different substrates—**1** and **3** (entries 1 and 3 in Table 2)—at 60 °C in DMF with no free base showed poor product yields. For example, it took 24 h in the absence of microwaves to yield **1s** in 60% yield, while **3s** was not detected even after 24 h (Table 2, entry 3). These results highlight the importance of microwaves in achieving rapid per-sulfation.

Table 1. Optimization of microwave-assisted sulfation of tetrahydroisoquinoline derivative **1**



1 **1s** (S=SO_3Na)

	Time (min)	Temperature (°C)	Modifications to conditions	HPLC yield (%)
1	20	100	No base	4.7
2	20	100	1 equiv Et_3N per OH grp	13.5
3	20	100	5 equiv Et_3N per OH grp	79.8
4	20	100	—[a]	80.0
5	10	100	1 equiv $SO_3{\cdot}Me_3N$ per OH grp	14.5
6	10	100	3 equiv $SO_3{\cdot}Me_3N$ per OH grp	46.1
7	10	100	—[a]	47.4
8	10	100	9 equiv $SO_3{\cdot}Me_3N$ per OH grp	79.5
9	10	100	12 equiv $SO_3{\cdot}Me_3N$ per OH grp	80.7
10	10	100	DMF as solvent	17.2
11	10	100	CH_3NO_2 as solvent	23.2
12	10	100	With 6 equiv $SO_3{\cdot}$py/OH grp and 10 equiv py/OH grp as base	82.3
13	10	40	—[a]	0
14	10	70	—[a]	18.2
15	10	120	—[a]	90.8
16	30	100	—[a]	91.2

[a] Reaction conditions here are as listed above with no additional modifications.

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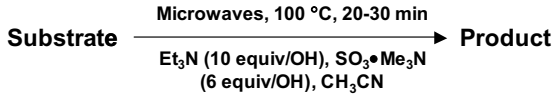
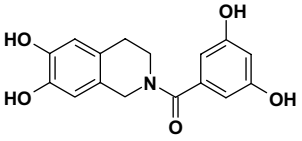
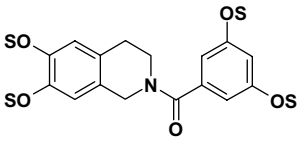
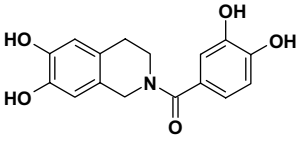
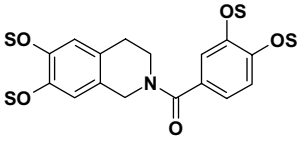
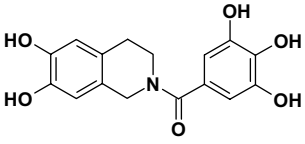
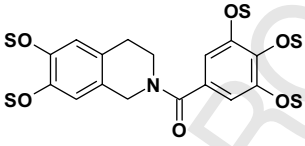
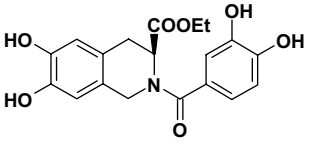
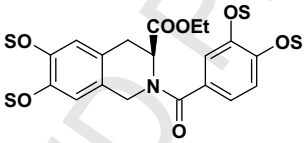
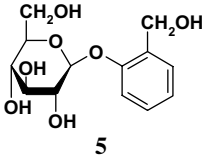
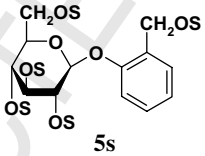
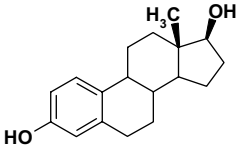
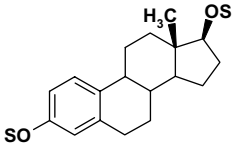
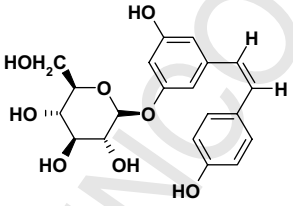
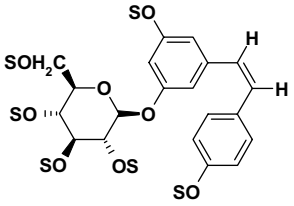
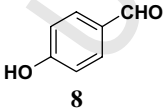
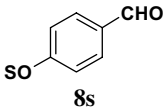
Table 2. Microwave-assisted sulfation of poly-hydroxyl substrates



	Substrate	Product[a]	Isolated yield (%)
1	 **1**	 **1s**	85
2	 **2**	 **2s**	87
3	 **3**	 **3s**	54[b]
4	 **4**	 **4s**	74
5	 **5**	 **5s**	84[c]
6	 **6**	 **6s**	94[c]
7	 **7**	 **7s**	72[d]
8	 **8**	 **8s**	97[c, e]

[a] S: SO_3Na.
[b] With 9 equiv of $SO_3{\cdot}Me_3N$/OH group.
[c] With $SO_3{\cdot}$py (6–9 equiv/OH group) and pyridine as base.
[d] Reaction conditions: 120 °C, 10 min, $SO_3{\cdot}$py (12 equiv/OH group) and pyridine as base.
[e] $S = SO_3{\cdot}pyH^+$.

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Having optimized the reaction conditions, we assessed whether the method works for a variety of different substrates. Per-sulfation of **2** proceeded smoothly in a manner identical to **1** (Table 2).[24] More importantly, per-sulfation of **3**, containing the crowded 3,4,5-trihydroxy moiety, was achieved under microwave conditions in an isolated yield of 54%, while the conventional procedure completely failed to give **3s**. Finally, microwave-assisted per-sulfation also works extremely well for substrates **5** through **8** containing one to six –OH groups. Interestingly, **5** and **6** gave a mixture of products with $SO_3{\cdot}Me_3N$, but yielded the per-sulfated products with $SO_3{\cdot}$py.

Several points make the microwave-assisted synthetic protocol particularly attractive. (A) The method appears to tolerate a range of functional groups including amide (Table 2, entries 1–4), ester (entry 4), aldehyde (entry 8) and double bond (entry 7). The relatively high isolated yields (~70–95%) in each case make the reaction especially suitable for library construction. (B) The methods work equally well for substrates containing one –OH group to those that contain six –OH groups. This is important because the small size of these molecules introduces considerable anion–anion repulsion as the number of sulfate groups increases. (C) The method applies equally well to alcoholic and phenolic –OH groups, especially with $SO_3{\cdot}$py complex. (D) The method provides high purity per-sulfated product that is readily isolated using an aqueous G10 filtration column. Typically, the purity of these highly water soluble, per-sulfated, small, organic molecules was found to be more than 95% using reverse polarity capillary electrophoresis (see Supplementary data). (E) The method is particularly suitable for quantitative isolation of small amounts (<10 mg) of the per-sulfated products, however could be linearly scaled up at least 20-fold without affecting the yields to a significant extent.

In summary, we have developed a rapid and high yielding microwave-based synthesis of variably functionalized, per-sulfated organic molecules. The protocol is expected to greatly facilitate the construction of a library of per-sulfated, small organic molecules for screening as glycosaminoglycan mimetics.

Acknowledgments

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Supplementary data

Preparation and characterization (NMR, mass, and capillary electrophoretic data) of the molecules prepared. Supplementary data associated with this article can be found, in the online version, at doi:10.1016/j.tetlet.2007.07.100.

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21. For a recent review on the impact of microwaves on drug discovery, see: Kappe, O. C.; Doris Dallinger, D. *Nat. Rev. Drug Discov.* **2006**, *51*, 51–63.
22. See Supplementary data for synthesis of starting materials.
23. RP-HPLC profile showed peaks from 4.3 to 6.0 min, in addition to one at 9.0 min. The peak at 4.3 was subsequently isolated after optimization of conditions and determined to be per-sulfated (**1s**). The peak at 9.0 min was identified as **1** by comparison with synthetically pure sample. Conversions (%) were determined by area normalization.
24. Representative procedure for per-sulfation: To a stirred solution of the poly-alcohol (20 mg, 0.066 mmol) in MeCN (1 mL) at rt, Et_3N (0.4 mL, 2.9 mmol) and $Me_3N{\cdot}SO_3$ (220 mg, 1.6 mmol) were added. The reaction vessel was sealed and micro-waved (CEM-discover microwave synthesizer) for 20 min at 100 °C. The reaction was repeated for four times and the reaction mixture was pooled for isolation of the product. The MeCN layer was decanted and pooled, while the residue was washed with MeCN (5 mL) and centrifuged. The combined MeCN layers were concentrated in vacuo. Water (5 mL) was added to the residue and stirred for 10 min. The water

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layer was concentrated to approximately 2 mL, loaded onto a Sephadex G10 column (~160 cm) and chromatographed using water as eluent. Fractions were combined based on RP-HPLC profiles, concentrated and re-loaded onto a SP Sephadex C25 column for sodium exchange. Appropriate fractions were pooled, concentrated in vacuo, and lyophilized to obtain a white powder. Spectral characteristics of the final sulfated compounds are as follows: **1s**: ^{1}H NMR (DMSO, 400 MHz) δ: 7.29–7.30 (m, 2H), 6.94–6.97 (m, 3H), 4.58 (s, 2H, isomer I), 4.48 (s, 2H, isomer II), 3.58 (s, 2H, isomer II), 3.50 (s, 2H, isomer I), 2.66 (br, 2H, isomer I and II); ESI (−ve) *m*/*z* Calcd for $C_{16}H_{11}NNa_4O_{17}S_4$ [(M−Na)$^-$] 685.86; found, 686.1. Compound **2s**: ^{1}H NMR (DMSO, 400 MHz) δ: 7.65 (d, $J = 2.4$ Hz, 1H), 7.57 (d, $J = 8.4$ Hz, 1H), 7.29 (s, 2H), 6.99 (dd, $J = 8.4$, 1.6 Hz, 1H), 4.54 (s, 2H), 3.70 (br, 2H), 2.69 (t, $J = 4.8$ Hz, 2H); ESI (−ve) *m*/*z* Calcd for $C_{16}H_{11}NNa_4O_{17}S_4$ [(M−Na)$^-$] 685.86; found, 686.0. Compound **3s**: ^{1}H NMR (DMSO, 400 MHz) δ: 7.37 (s, 2H), 7.29 (s, 2H), 4.54 (s, 2H), 3.53 (s, 2H), 2.68 (s, 2H); ESI (−ve) *m*/*z* Calcd for $C_{16}H_{10}NNa_5O_{21}S_5$ [(M−Na)$^-$] 803.79; found, 804.1. Compound **4s**: ^{1}H NMR (DMSO, 400 MHz) δ: 6.96–7.70 (m, 5H), 4.83–5.16 (m, 1H isomers I–III), 4.18–4.44 (m, 2H, isomers I–III), 3.55–3.61 (m, 2H, isomers I–III), 2.99–3.13 (m, 2H, isomers I–III), 1.08–1.16 (m, 3H, isomers I–III); ESI (−ve) *m*/*z* Calcd for $C_{19}H_{15}NNa_4O_{19}S_4$ [(M−Na)$^-$] 757.88; found, 757.5. Compound **5s**: ^{1}H NMR (DMSO, 400 MHz) δ: 7.34 (d, 1H, $J = 8.0$ Hz), 7.16 (t, 1H, $J = 8.0$ Hz), 6.95 (t, 1H, $J = 8.0$ Hz), 6.89 (d, 1H, $J = 8.0$ Hz), 5.24 (d, 1H, $J = 8.0$ Hz), 4.86 (s,2H), 4.62 (d, 2H, $J = 4.0$ Hz), 4.35 (m, 1H), 4.21–4.27 (m, 1H), 3.93–3.99 (m, 1H), 3.75 (m, 1H); ESI (−ve) *m*/*z* Calcd for $C_{16}H_{10}NNa_5O_{21}S_5$ [(M−Na)$^-$] 772.81; found, 772.7. Compound **6s**: ^{1}H NMR (DMSO, 400 MHz) δ: 7.1 (d, $J = 8.4$ Hz, 1H), 6.81–6.83 (m, 2H), 4.02 (t, $J = 8.0$ Hz, 1H), 2.71–2.74 (m, 2H), 2.21–2.25 (m, 1H), 2.08–2.12 (m, 1H), 1.87–2.00 (m, 2H), 1.75–1.78 (m, 1H), 1.48–1.60 (m, 2H), 1.09–1.34 (m, 6H), 0.66 (s, 3H); ESI (−ve) *m*/*z* Calcd for $C_{18}H_{22}Na_2O_8S_2$ [(M−Na)$^-$] 453.07; found, 453.1. Compound **7s**: ^{1}H NMR (DMSO, 400 MHz) δ: 7.56 (s, 1H), 7.54 (s, 1H), 6.99–7.10 (m, 7H), 6.61 (s, 1H), 5.36 (s, 1H), 4.63 (s, 1H), 4.59 (s, 1H), 4.39 (s, 1H), 4.14–4.18 (m, 1H), 4.08 (d, $J = 8.8$ Hz, 1H) 3.81 (t, $J = 10$ Hz, 1H); ESI (−ve) *m*/*z* Calcd for $C_{20}H_{16}Na_6O_{26}S_6$ [(M−Na)$^-$] 978.77; found, 979.0. Compound **8s**: ^{1}H NMR (DMSO, 400 MHz) δ: 9.76 (s, 1H), 8.94–8.96 (m, 2H), 8.62–8.68 (m, 1H), 8.09–8.14 (m, 2H), 7.72–7.75 (m, 2H), 6.92–6.95 (m, 2H); ESI (−ve) *m*/*z* Calcd for $C_{12}H_{11}NO_5S$ [(M−pyH$^+$)$^-$] 200.99; found, 200.8.

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