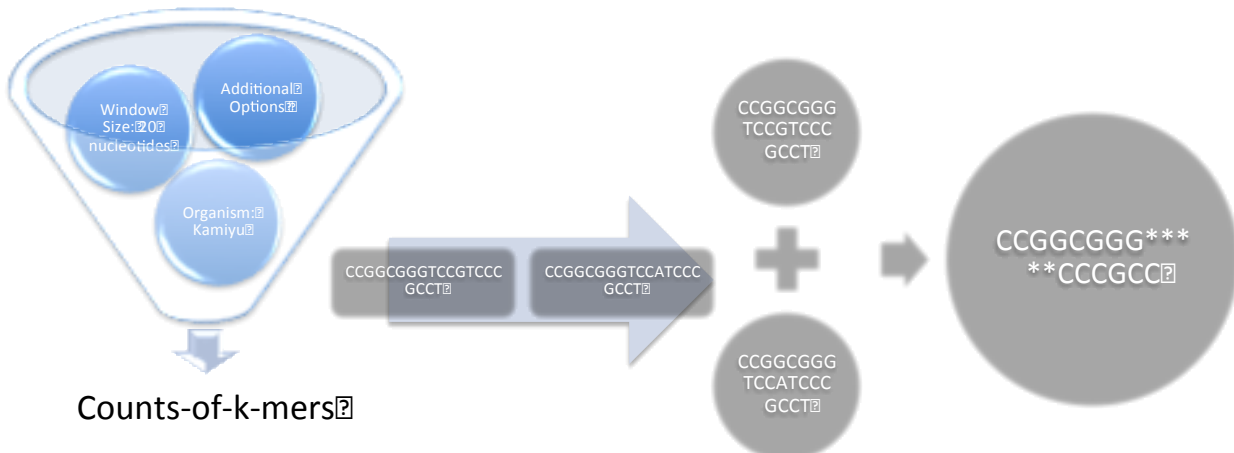


## Introduction

Let's start the expedition. Our group explored the realm of short dispersed repeats, and attempted to derive meaning from their functionality. Short dispersed repeats are approximately twenty to two hundred nucleotides long and occur in various genomes of both bacteria and viruses. For my portion of the project I decided to look at short dispersed repeats that occur in mycobacteriophages or viruses that infect mycobacteria such as Mycobacterium Tuberculosis and Mycobacterium Smegmatis which is a lab strain I have previously worked with. Since I was involved in Phages Discovered lab in the beginning of my sophomore year I have grown an interest in discovering what these viruses are capable of doing. Originally I was at a loss for where my starting point would be, and given my disdain for reading scientific papers I decided (rightly or wrongly) that I would go into the genome of mycobacteriophages and identify possible short dispersed repeats.

## Methods



So initially I wanted to find something novel so my first choice was to look for twenty nucleotide repeats in mycobacteriophages using the counts-of-k-mers function in BioBike, which is a visual interactive programming framework where students can analyze genomic sequences. The function counts-of-k-mers works by going through the genome of a specified organism and identifying repeats based on the sequence size the user designates. For example if I (the user) wanted to look for twenty nucleotide repeats the function will go through the genome and identify repetitive twenty nucleotide sequences, and generate a results pair that will display the actual repetitive sequences with the number of times the sequence repeats in the genome. In order to test out whether this mechanism would be useful I picked a random mycobacteriophage and ran the counts-of-k-mer and defined it so that it would identify twenty nucleotide repetitive sequences. The mycobacteriophage I happened to choose was a sub-cluster B3 bacteriophage named Kamiyu. Using this function I found the short dispersed sequence "GGCGGGTCCGTCCTCCGCC", which occurred three times in mycobacteriophage Kamiyu. I proceeded to use another function called matches-by-pattern, this function allows the user to input a sequence and an organism to look into for that sequence in the organism by matching its patterns in that target organism. I primarily used this function to get the coordinates of the sequence and identify

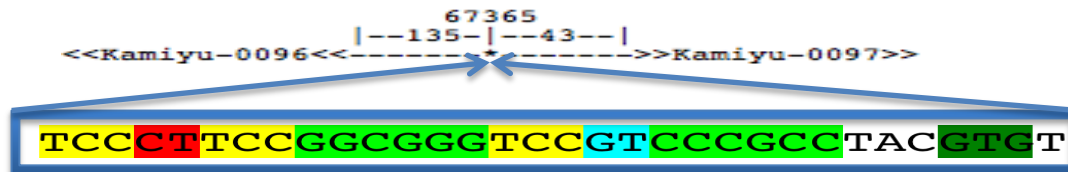
locations of these sequences in Kamiyu. I was able to find additional sequences that were not identified in count-of-k-mers, and find interesting location specific tendencies. Although the sequence itself may not seem particularly remarkable the location of the repeats struck me as they mostly occurred at the tail end of the mycobacteriophage's sequence. I went through by hand and lined up the sequences on top of each other with the coordinates next to each one and looked for motifs, which I found as horribly inefficient and there was actually a function that does that called motif in but the time had already passed so I moved on. When I did this I saw a hairpin sequences that occurred with a five-nucleotide long loops.

Key	KAMIYU			
Green: Palindrome (Suspect RNA Hairpin)	17587	17618	CCGCTTCCGGCGGGTCCGTCCCGCCTGC	GTGA F
Yellow: Conserved short repeat of three nucleotides	60845	60876	TCCCTTCCGGCGGGTCCGTCCCGCCTTC	ATGA F
Red: Partially Conserved duplicate	67236	67267	CCTTCCCGCGGGTCCGTCCCGCCTACACC	F
Blue: Partially Conserved duplicate	67398	67365	TCCCTTCCGGCGGGTCCGTCCCGCCTAC	GTG B
	68276	68243	TCCCTTCCGGCGGGTCCGTCCCGCCTCC	GTGA B

For my next step I wanted to see if this sequence occurred in other bacteriophages in similar locations and in order to do this I used matches-by-pattern but looked at another phage in the same cluster as Kamiyu, called Athena and was able to see a similar pattern in location and sequence although insertions caused shifts further downstream.

Key	ATHENA			
Green: Palindrome (Suspect RNA Hairpin)	18381	18412	CCGCTTCCGGCGGGTCCGTCCCGCCTTC	GTGA F
Yellow: Conserved short repeat of three nucleotides	45810	45841	TCCCTTCCGGCGGGTCCGTCCCGCCTTCGA	TC F
Red: Partially Conserved duplicate	61621	61652	TCCCTTCCGGCGGGTCCGTCCCGCCTTC	ATGA F
Blue: Partially Conserved duplicate	68011	68042	CCTTCCCGCGGGTCCGTCCCGCCTACACC	F
	68173	68140	TCCCTTCCGGCGGGTCCGTCCCGCCTAC	GTG B
	69052	69019	TCCCTTCCGGCGGGTCCGTCCCGCCTCC	GTGA B

As to continue this process more efficiently I used the for-each loop function to map over the all mycobacteriophages list, which contained all the mycobacteriophage contained in the BioBike database. The for-each loop works by extracting each the name of each mycobacteriophages from the list of mycobacteriophages and inputting the phage into the body of the loop which was define as matches by pattern in this case and giving back similar results as above, but for each phage it actually found the sequence in on this the mycobacteriophages list. I only took phages where the sequence occurred in the same amount or more than Kamiyu, and in all cases those phages were in subcluster B3 phages that are available through biobike.



Actual nucleotide sequence is contained in the box above and the asterisk represents its context in terms of the first letter in the nucleotide sequence. The flanking regions are the genes it is in-between and the numbers to the left and right represent how far it is from the end or beginning of the reading frame for those particular proteins.

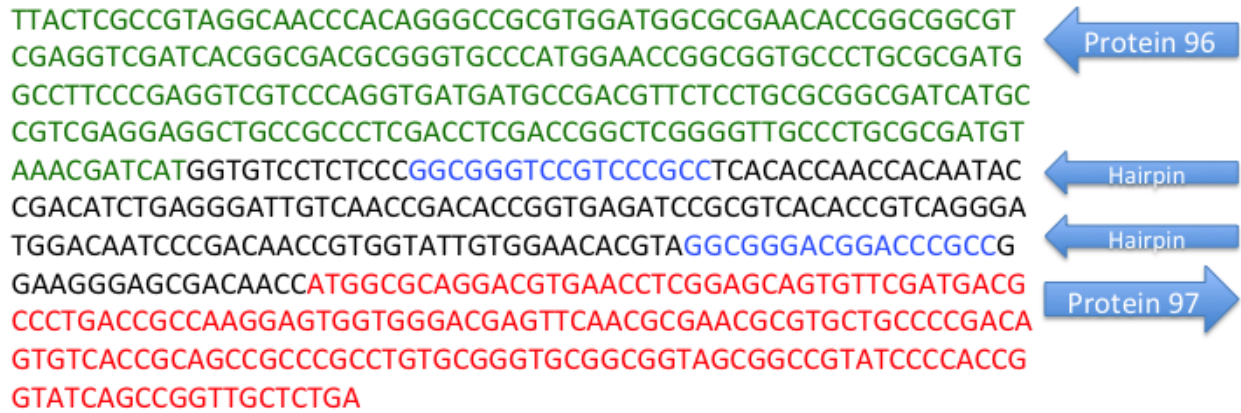


Filtered out only divergent intergenic sequences then re-filtered for divergent intergenic sequences with two palindromes

My next step was to look at the context of each of these sequences by taking the coordinates and their phage name and inserting it into the context-of function. This function works to tell the user where the sequence is in relation to the coding proteins of the genome. And, this occupied the bulk of my time but I ultimately did not use it thoroughly in my data generating given that it would return faulty matches that did not actually exist in the genomes. Although I still continued to use context of my extracting the sequence coordinates and mycobacteriophages names from the sequence similar to function. Many of the sequences were in genes that were transcribed parallel to on another, but some did occurring in genes that were divergent to one another and this is where my attention ended up shifting.

## Results

I filter for only the divergent sequences and from there I filtered for the two divergent sequences that occurred in the same intergenic sequence for all the phages and in Kamiyu that happened to be protein 96 and protein 97 although that was not the case for the other phages, although for these two sequences the distance between any two protein there were present in was generally conserved.



Region with blue lettering represents the dual palindromic sequences in the intergenic region between Kamiyu Protein 96 and Protein 97

Interestingly enough there were not convergent sequences where their palindromes were located which may be a indicator to what their functions will be.

#### Discussion

Possibly that most difficult portion of this project was determining what the purpose of my actual sequences were. Just looking at the region with their palindromes I looked at the domain of the upstream and downstream proteins to see if there were any prediction functions and I was able to find a coiled-coil structure upstream in Kamiyu and a membrane protein structure downstream from the sequence in Kamiyu. Another thing that I did not have time to do additional research on but observed recently was that although the identified sequences occur in genes transcribed parallel to the sequence there was an interesting overrepresentation of that sequence in divergent regions

My view of these sequences is that they may serve as genetic switches similar to *cro* and *cl* of lambda phage or directional switches that control protein production that may be related to their lytic or lysogenic phase of the phage.

## Appendix

### Motifs in Original Phage Cluster

```
KAMIYU
17587 17618 CCGCTTCGGGCGGGTCCGTCCCGCTTCGTTAA F
60845 60876 TCCTCTCCGGCGGGTCCGTCCCGCTTCGTTAA F
67236 67267 CCTCTCCGGCGGGTCCGTCCCGCTTCACACCF F
67398 67365 TCCCTTCGGGCGGGTCCGTCCCGCTTACGTT B
68276 68243 TCCCTTCGGGCGGGTCCGTCCCGCTTCGTTAA B

ATHENA
18381 18412 CCGCTTCGGGCGGGTCCGTCCCGCTTCGTTAA F
45810 45841 TCCCTTCGGGCGGGTCCGTCCCGCTTCGTTAA F
61621 61652 TCCTCTCCGGCGGGTCCGTCCCGCTTCGTTAA F
68011 68042 CCTCTCCGGCGGGTCCGTCCCGCTTCACACCF F
68173 68140 TCCCTTCGGGCGGGTCCGTCCCGCTTACGTT B
69052 69019 TCCCTTCGGGCGGGTCCGTCCCGCTTCGTTAA B

YAHALOM
17628 17659 CCGCTTCGGGCGGGTCCGTCCCGCTTCGTTAA F
45074 45105 TCCCTTCGGGCGGGTCCGTCCCGCTTCGTTAA F
60695 60726 TCCTCTCCGGCGGGTCCGTCCCGCTTCGTTAA F
67085 67116 CCTCTCCGGCGGGTCCGTCCCGCTTCACACCF F
67247 67214 TCCCTTCGGGCGGGTCCGTCCCGCTTACGTT B
68125 68092 TCCCTTCGGGCGGGTCCGTCCCGCTTCGTTAA B

GADJET
17622 17653 CCGCTTCGGGCGGGTCCGTCCCGCTTCGTTAA F
45080 45111 CCTCTCCGGCGGGTCCGTCCCGCTTCGTTAA F
61248 61279 TCCTCTCCGGCGGGTCCGTCCCGCTTCGTTAA F
62890 62921 TCCCTTCGGGCGGGTCCGTCCCGCTTCACACCF F
66542 66573 CCTCTCCGGCGGGTCCGTCCCGCTTCACACCF F
66704 66671 TCCCTTCGGGCGGGTCCGTCCCGCTTACGTT B
67592 67559 TCCCTTCGGGCGGGTCCGTCCCGCTTACGTT B

PHAEDRUS
17534 17565 CCGCTTCGGGCGGGTCCGTCCCGCTTCGTTAA F
44985 45016 TCCCTTCGGGCGGGTCCGTCCCGCTTCGTTAA F
66693 66724 CCTCTCCGGCGGGTCCGTCCCGCTTCACACCF F
66855 66822 TCCCTTCGGGCGGGTCCGTCCCGCTTACGTT B
67733 67700 TCCCTTCGGGCGGGTCCGTCCCGCTTCGTTAA B

PHYLER
18353 18384 CCGCTTCGGGCGGGTCCGTCCCGCTTCGTTAA F
45802 45833 TCCCTTCGGGCGGGTCCGTCCCGCTTCGTTAA F
61598 61629 TCCTCTCCGGCGGGTCCGTCCCGCTTCGTTAA F
67982 68013 CCTCTCCGGCGGGTCCGTCCCGCTTCACACCF F
68144 68111 TCCCTTCGGGCGGGTCCGTCCCGCTTACGTT B
69021 68988 TCCCTTCGGGCGGGTCCGTCCCGCTTACGTT B

DAISY
17639 17670 CCGCTTCGGGCGGGTCCGTCCCGCTTCGTTAA F
45087 45118 TCCCTTCGGGCGGGTCCGTCCCGCTTCGTTAA F
60448 60479 TCCTCTCCGGCGGGTCCGTCCCGCTTCGTTAA F
66838 66869 CCTCTCCGGCGGGTCCGTCCCGCTTCACACCF F
67000 66967 TCCCTTCGGGCGGGTCCGTCCCGCTTACGTT B
67888 67855 TCCCTTCGGGCGGGTCCGTCCCGCTTCGTTAA B

PIPERFISH
18500 18531 CCGCTTCGGGCGGGTCCGTCCCGCTTCGTTAA F
61994 62025 TCCTCTCCGGCGGGTCCGTCCCGCTTCGTTAA F
63634 63665 TCCGTTCGGGCGGGTCCGTCCCGCTGCACCT F
67662 67693 CCTTTCCGGCGGGTCCGTCCCGCTTCACACCF F
67824 67791 TCCCTTCGGGCGGGTCCGTCCCGCTTACGTT B
68702 68669 TCCCTTCGGGCGGGTCCGTCCCGCTTCGTTAA B

AKOMA
17616 17647 CCGCTTCGGGCGGGTCCGTCCCGCTTCGTTAA F
45063 45094 TCCCTTCGGGCGGGTCCGTCCCGCTTCGTTAA F
60886 60917 TCCTCTCCGGCGGGTCCGTCCCGCTTCGTTAA F
67315 67346 CCTCTCCGGCGGGTCCGTCCCGCTTCACACCF F
67477 67444 TCCCTTCGGGCGGGTCCGTCCCGCTTACGTT B
68354 68321 TCCCTTCGGGCGGGTCCGTCCCGCTTACGTT B
```

### Dual Palindrome Sites

```
((D Akoma.Akoma-0097 Akoma.Akoma-0098 160 18 B)
(D Athena.Athena-0096 Athena.Athena-0097 160 18 B)
(D Daisy.Daisy-0095 Daisy.Daisy-0096 160 18 B)
(D Gadget.Gadget-0093 Gadget.Gadget-0094 160 18 B)
(D Kamiyu.Kamiyu-0096 Kamiyu.Kamiyu-0097 160 18 B)
(D Phaedrus.PHAEDRUS 93 Phaedrus.PHAEDRUS 94 160 18 B)
(D Phlyer.PHLYER 98 Phlyer.PHLYER 99 160 18 B)
(D Pipefish.Pipefishp97 Pipefish.Pipefishp98 160 18 B)
(D Yahalom.Yahalom-0095 Yahalom.Yahalom-0096 160 18 B))
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```
((D Akoma.Akoma-0097 Akoma.Akoma-0098 30 148 B)
(D Athena.Athena-0096 Athena.Athena-0097 30 148 B)
(D Daisy.Daisy-0095 Daisy.Daisy-0096 30 148 B)
(D Gadget.Gadget-0093 Gadget.Gadget-0094 30 148 B)
(D Kamiyu.Kamiyu-0096 Kamiyu.Kamiyu-0097 30 148 B)
(D Phaedrus.PHAEDRUS 93 Phaedrus.PHAEDRUS 94 30 148 B)
(D Phlyer.PHLYER 98 Phlyer.PHLYER 99 30 148 B)
(D Pipefish.Pipefishp97 Pipefish.Pipefishp98 30 148 B)
(D Yahalom.Yahalom-0095 Yahalom.Yahalom-0096 30 148 B))
```